a-HYDROXYLATED DERIVATIVES OF ANTITUMOUR DIMETHYLTRIAZENES

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The prophetic suggestion 1 that metabolic α -hydroxylation could play a critical role in the carcinogenic activity of dimethylnitrosamine has been sustained by the recent development of synthetic routes to α -acetoxydimethylnitrosamine (1).² Unlike dimethylnitrosamine the acetoxy derivative is profoundly mutagenic without prior metabolic activation.³⁻⁵ $Chemical$ studies with a range of a-acetoxymethylnitrosamines corroborate the theory that such compounds are non-enzymatically deacetylated to unstable hydroxymethyl derivatives which decompose to afford alkylating species.^{2,6,7}

A similar oxidative activation process has been proposed to explain the fact that l-aryl-3,3-dimethyltriazenes (3) although inactive in vitro develop antitumour properties in vivo or after incubation with liver fractions. 8.9 In these cases alkylating monomethyltriazenes (4) are implicated as the cytotoxic species. Although the related α -hydroxymethyl intermediates (2) and (51 have hitherto been regarded as only transient species we have achieved a synthesis of the latter type simply by reacting aryldiazonium salts with a formaldehyde-methylamine mixture.

In a typical synthesis methyl p -aminobenzoate (4.0 g), diazotised as normal in 1.5Nhydrochloric acid (60 ml) with sodium nitrite (1 mol. equiv.), was added (15 min.) to a pre-mixed solution of 40% aqueous formaldehyde (60 ml) and 25% aqueous methylamine (6 ml) at -5° . The white hydroxymethyltriazene (5a) was collected, washed with water, air-dried and crystallised from benzene. Physical characteristics of (5a) and related derivatives are listed in the Table.

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The $\frac{1}{1}$ H n.m.r. spectrum of (5a) recorded in CDCl₃ showed singlets at 63.3 (N-CH₃) and 3.95 $(O-CH_3)$, a doublet at δ 5.3 (H_2) which collapsed to a singlet with D_2O , and a typical aromatic AA'BB' pattern centred at 67.8. The i.r. spectrum of (5a) showed significant peaks at 3440 (OH) and 1695 cm^{-1} (CO). More informative was the mass spectrum which showed a small molecular ion at m/e 223 (Relative Intensity \sim 1% at 70 eV) and significant peaks at m/e 193 (10%; M - CH_nO), m/e 163 (20%) and 151 (100%). The peak at 163 corresponds to the aryldiazonium ion fragment and that at 151 to the radical ion of methyl p-aminohenzoate. The appearance of abundant arylamine radical ion peaks in the mass spectra of all the hydroxymethyltriazenes is significant since only those triazenes which can fragment to monoalkyltriazenes normally afford such ions. ¹⁰ They are absent, for example, in the spectra of the dimethyltriazenes (3; $X = p$ -CO₂Me or P-Ac) where the base peaks are the appropriate aryldiazonium ions, and feature to only a minor extent in the spectrum of the benzoate ester of (5a).

Table. a-Hydroxymethyltriazenes (5) from diazonium salts and formaldehyde-methylamine

* All compounds gave satisfactory CHN analyses.

Prepared from (5a) with benzoyl chloride in pyridine at 25[°].

The hydroxymethyltriazene (5a) afforded a crystalline henzoate ester but decomposed in acetic anhydride or acetyl chloride-pyridine. Although stable in non-polar solvents the hydroxymethyltriazene (5a) rapidly decomposed by a first order process ($t_{1/2} = 22$ min.) in 0.5 M phosphate buffer (pH 7.5) at 25[°]. When the decomposition of (5a) was followed spectroscopically (λ max. 313 nm) the final spectrum was identical with that of methyl p -aminobenzoate and may be envisaged as proceeding by initial loss of formaldehyde according to Scheme 1.

Scheme 1

However, decomposition of the bensoate ester of (Sal which also ultimately forms methyl p-aminobenzoate occurs at a rate faster ($t_{\frac{1}{2}}$ = 26.5 min.) than can be explained simply by invoking an initial hydrolysis of the benzoate group. In this case we propose an alternative route (Scheme 2) via an imminium intermediate (6) similar to that suggested to explain the anomalous stabilities of some esters of structurally related α -hydroxylated nitrosamines.

Scheme 2

Successful syntheses of hydroxymethyltriazenes (5) were only achieved from diazonium salts bearing - **M** substituents in the p-position of the aryl group; only unstable oils were formed from p -toluidine or p -chloroaniline. Attempted synthesis of a 'masked' hydroxymethyltriazene the lactone (7) - from diazotised methyl anthranilate led to the formation of a high yield of the triazinone (8) presumably by cyclisation of a monomethyltriazene (4; $X = \sigma - \text{CO}_2$ Me) liberated by decomposition of the precursor hydroxymethyltriazene (5; $X = 0-C0$ ₂Me).¹¹

The hydroxymethyltriaxene (5a) has pronounced antitumour properties giving a maximum percentage increase in survival time (IST) of 120% (5 daily doses of 5 mg/Kg)in mice innoculated with the TLX5 lymphoma. Comparable IST values for the related di- and mono-methyltriasenes (3 and 4; $X = p-CO_2$ Me) are 58% (at 40 mg/Kg) and 87% (at 5 mg/Kg) respectively. Moreover, the ethyl ester (5b) is clearly an activated triazene since it inhibits the TLX5 lymphoma in vitro whereas its dimethyltriazene counterpart (3; $X = p - \text{CO}_2 E$) is inactive.

The close parallel between the chemical and biological properties of dimethylnitrosamine and aryldimethyltriazenes is further emphasised by the strong mutagenic activity of the hydroxymethyltriazenes (5a) and (5c) towards *Salmonella typhimurium* TA 100 and TA 1535; the corresponding dimethyltriazenes (3; $X = p - CO_2$ Me or $p - AC$) are non-mutagenic unless activated by liver fractions.

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References:

- 1. H. Druckrey, R. Preussmann, D. Schmänl and M. Müller, *Naturwissenschaften*, 48, 134 (1961).
- *2.* P.P. Roller, D.R. Shimp and L.K. Keefer, *Tetrahedron Letters, 2065* (1975).
- *3. O.G.* Fahmy, M.J. Fahmy and **M.** Wiessler, *Biochem. PharmacoZ., 24,* 1145 (1975).
- 4. O.G. Fahmy and M.J. Fahmy, Cancer *Res., 35, 3780* (1975).
- *5. M.* Okada, E. Suzuki, T. Anjyo and M. Mochizuki, **Gum,** *66, 457 (1975); Chem. Abs., 83,* 189091h (1975).
- 6. P.L. Skipper, S.R. Tannenbaum, J.E. Baldwin and A. Scott, *Tetrahedron Letters, 4269 (1977).*
- *7.* J.E. Baldwin, A. Scott, S.E. Branz, S.R. Tannenbaum and L. Green, J. **Org.** *Chem., 43, 2427 (1978).*
- *8.* R.C.S. Audette, T.A. Connors, H.G. Mandel, K. Merai and W.C.J. Ross, *Biochem. Pharmacol., 22,* 1855 (1973).
- 9. T.A. Connors, P.M. Goddard, K. Merai, W.C.J. Ross and D.E.V. Wilnan, *Biochem. Pharmacol., 25, 241 (1976).*
- **10.** G.F. Kolar in "Mass Spectrometry in Biochemistry and Medicine"; Frigerio and Castagnoli Eds., Raven Press, New York, p.267 (1974).
- 11. R.J. LeBlanc and K. Vaughan, **Can.** *J. Chem., 50, 2544 (1972).*

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